

Compact 3U as a novel lighting source for the propagation of some horticultural plants

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Abstract

A novel lighting system (Compact 3U) was successfully applied to the micropropagation of some horticultural plants. *Cymbidium* 'Tim Hot', *Lilium longiflorum* and *Fragaria vesca* cv. 'My Da' shoots were used for this study. To compare *in vitro* growth of plantlets placed under Neon and Compact 3U lighting systems, *Fragaria vesca* cv. 'My Da' shoots were cultured on ½ MS medium supplemented with 1 g⁻¹ activated charcoal, 30 g⁻¹ sucrose and 8 g⁻¹ agar under two lighting sources at 45 μmolm⁻²s⁻¹. After three weeks of culture, the shoot and root length, leaf area and fresh weight of strawberry plantlets under Compact 3U system were significantly higher than those grown under Neon system. To clarify the effect of irradiance of Compact 3U system on the development of plantlets, *Cymbidium* 'Tim Hot' shoots were cultured on MS medium supplemented with 0.5 mg⁻¹ NAA, 1 g⁻¹ activated charcoal, 100% coconut water, 25 g⁻¹ sucrose and 8 g⁻¹ agar, *Lilium longiflorum* and *Fragaria vesca* cv. 'My Da' shoots were cultured on ½ MS medium supplemented with 1 g⁻¹ activated charcoal, 30 g⁻¹ sucrose and 8 g⁻¹ agar at different irradiances: (1) Neon at 45 μmolm⁻²s⁻¹ (control), and Compact 3U at: (2) 45 μmolm⁻²s⁻¹, (3) 60 μmolm⁻²s⁻¹, and (4) 75 μmolm⁻²s⁻¹. The results showed that plantlets of the three genera adapted differently to irradiances and lighting sources, but in all, the growth of plantlets were better under the Compact 3U system. Furthermore, *ex vitro* plantlets derived from Compact 3U system also developed better than those from Neon system.

Key words: Compact 3U, Neon, *Cymbidium* 'Tim Hot', *Lilium longiflorum*, *Fragaria vesca* cv. 'My Da'

Introduction

Now-a-days, *in vitro* multiplication is a primary method to rapidly mass-produce horticultural plants. The demand for high quality planting material has been increasing quickly worldwide for reforestation, foods/forage production, urban/indoor horticulture and global environment protection (Kozai *et al.*, 1992). In many cases, since micropropagation gave some superior transplant qualities to seedling production and conventional vegetative production, billions of micropropagated plantlets were produced annually world-wide (Debergh and Zimmerman, 1990). Tissue culture has been carried out in more than 600 companies all over the world. However, the widespread use of micropropagation for major crops in agriculture and horticulture was restricted because of its relatively high production costs caused by high labour cost (Kozai *et al.*, 1992), especially electrical energy consumption.

Control of plantlet growth and morphology is important in micropropagation to obtain high plantlet quality at different growth and developmental stages, and to save labour by automation or robotics (Miyashita, 1995). Many of the growth and morphological characteristics of plants *in* and *ex vitro* are influenced by environmental factors, such as light (quality, intensity, duration and direction), temperature, gaseous composition (CO₂, O₂, H₂O and C₂H₄), and medium composition (Schwabe, 1963; Kozai *et al.*, 1992). Light quality had a significant influence on the growth and morphology of plants *in* and *ex vitro* (Warrington and Michell, 1976; Morgan and Smith, 1981; Smith, 1982; Tibbitts *et al.*, 1983; Mortensen and Stromme, 1987; Economou and Read, 1987; Agrawal, 1992). The total quantity of light that a plant received during illumination directly

affected photosynthesis as well as plant growth and yield (Kim and Kozai, 2000). Hence, many lighting systems that effectively used electrical energy in the multiplication of horticultural plants have been studied intensively such as fluorescent, incandescent, luminescent (Sodium high pressure) lighting systems, and recently, light-emitting diode (LED) lighting source. However, tissue cultured plants are almost invariably grown under fluorescent illumination (Collin *et al.*, 1988), especially under cool white fluorescent lamps with a high proportion of its output in the blue and red regions (Hart, 1988).

Previous studies had been done on the effect of light quality and intensity of different lighting sources on the growth and morphology of *in* and *ex vitro* plantlets (Seabrook, 1987; Hayashi *et al.*, 1992; Iwanami *et al.*, 1992; Kozai *et al.*, 1992; Kirdmanee *et al.*, 1993; Gabarkiewicz *et al.*, 1997; Wulster and Janes, 1997; Maas and Bakx, 1997; Kunneman and Ruesink, 1997; Moe, 1997; Faust and Heins, 1997; Murakami *et al.*, 1997; Gabryszewska and Rudnicki, 1997; Walz and Horn, 1997; Miyashita *et al.*, 1997; Nhut, 2002). In our study, Compact 3U lamps were used as a promising lighting source for propagating some horticultural plants such as *Cymbidium*, *Lilium* and strawberry. These plants are highly valuable economic crops in Vietnam as well as all over the world. In this report, we focused on the effects of two different lighting sources (Neon and Compact 3U) as well as some different intensities of Compact 3U lamps (45, 60, and 75 μmolm⁻²s⁻¹, respectively) on the growth and morphology of these *in vitro* plantlets, and Neon lamp (with cool white emission) as a control system.

Compact 3U lamp (Fig. 1), which saves 80% electrical energy

as compared to incandescent lamps, has a compact size, long life (>6.000 h), and reaches one-fifth the brightness of conventional incandescent lamps. Hence, plant production cost could be decreased.

Materials and methods

Plant materials and culture media: *Cymbidium* 'Tim Hot' shoots (4 cm length), derived from protocorm-like bodies (PLBs) cultured on MS (Murashige and Skoog, 1962) medium containing 0.5 mg l⁻¹ α -naphthaleneacetic acid (NAA), 2 mg l⁻¹ 6-benzyladenine (BA), 1 g l⁻¹ activated charcoal (AC), 20% coconut water (CW), 30 g l⁻¹ sucrose and 8 g l⁻¹ agar (Haiphong Co., Vietnam), were cultured on MS medium containing 0.5 mg l⁻¹ NAA, 1 g l⁻¹ AC, 10% CW, 25 g l⁻¹ sucrose and 8 g l⁻¹ agar.

Lilium longiflorum bulb scales, derived from *in vitro* bulblets cultured on MS medium containing 0.2 - 0.5 mg l⁻¹ BA, 30 g l⁻¹ sucrose and 8 g l⁻¹ agar, were cultured on ½ MS medium supplemented with 1 g l⁻¹ AC, 30 g l⁻¹ sucrose and 8 g l⁻¹ agar.

Fragaria vesca cv. 'My Da' shoots (1.5 cm), derived from meristems cultured on MS medium containing vitamin B₅, 0.2 mg l⁻¹ BA, 30 g l⁻¹ sucrose and 8 g l⁻¹ agar, were cultured on ½ MS medium supplemented with 1 g l⁻¹ AC, 30 g l⁻¹ sucrose and 8 g l⁻¹ agar.

For all experiments, explants were cultured in vessels (500 ml) containing 60 ml medium. pH of media was adjusted to 5.7 before autoclaving at 121°C, 1 atm for 40 min.

Lighting systems: Cool white fluorescent lamps (Neon tubes) (40 W each; Rang Dong Light source and Vacuum Flask Co., Vietnam, FL-40W/T10) and warm white fluorescent lamps (Compact 3U lamps) (18 W each; Rang Dong Light source and Vacuum Flask Co., Vietnam, CFH-3U18W) were used as lighting sources in each experiment.

Irradiances were 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ under Neon light or 45, 60, 75 $\mu\text{molm}^{-2}\text{s}^{-1}$ for Compact 3U system according to each experiment. Photosynthetic photon flux density (PPFD) was measured with

an illumination meter (Tokyo photoelectric Co., LTD., Japan, ANA-F11) on the empty culture shelf.

Experimental designs

Effect of Compact 3U lighting source on the *in vitro* development of *Fragaria vesca* cv. 'My Da' plantlets: Five strawberry shoots were cultured in each culture vessel, and ten vessels were placed on the shelf in one row under the Compact 3U lighting system with three lamps per shelf, arranged in one row. Ten other vessels were placed in one row on another shelf under the Neon lighting system at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ (the control lighting system). After three weeks of culture, some morphological parameters (plant height and fresh weight, root length, leaf area) were recorded and the *in vitro* plantlets were transplanted to the greenhouse.

Effect of different irradiances of Compact 3U on the *in vitro* development of *Cymbidium*, *Lilium* and strawberry plantlets: Each vessel contained five shoots of each plant (*Cymbidium*, *Lilium* and strawberry). There were four shelves (ten vessels per shelf) with different irradiances: three shelves with the Compact 3U lighting system at either 45, 60 or 75 $\mu\text{molm}^{-2}\text{s}^{-1}$, and the remaining shelf with the Neon lighting system at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$. Some morphological parameters of strawberry plantlets (plant height, leaf area, root length, and plant fresh weight) were recorded after three weeks of culture, of *Cymbidium* (plant height, root length, leaf area, number of newly formed roots, number of bulbs, bulb diameter, bulb cluster fresh weight and bulb fresh weight) and of *Lilium* (plant height, root length, leaf width, number of roots and plant fresh weight) were recorded after six weeks of culture.

The process to set up the Compact 3U and Neon lighting sources for studying *in vitro* development of some horticultural plants is depicted in Fig. 2.

The *in vitro* plantlets were thereafter transplanted to greenhouse. This subsequent stage of development of *Lilium* and *Cymbidium* plantlets were placed under 6h/day supplemental Compact 3U lighting source. After one and a half months of culture, plant

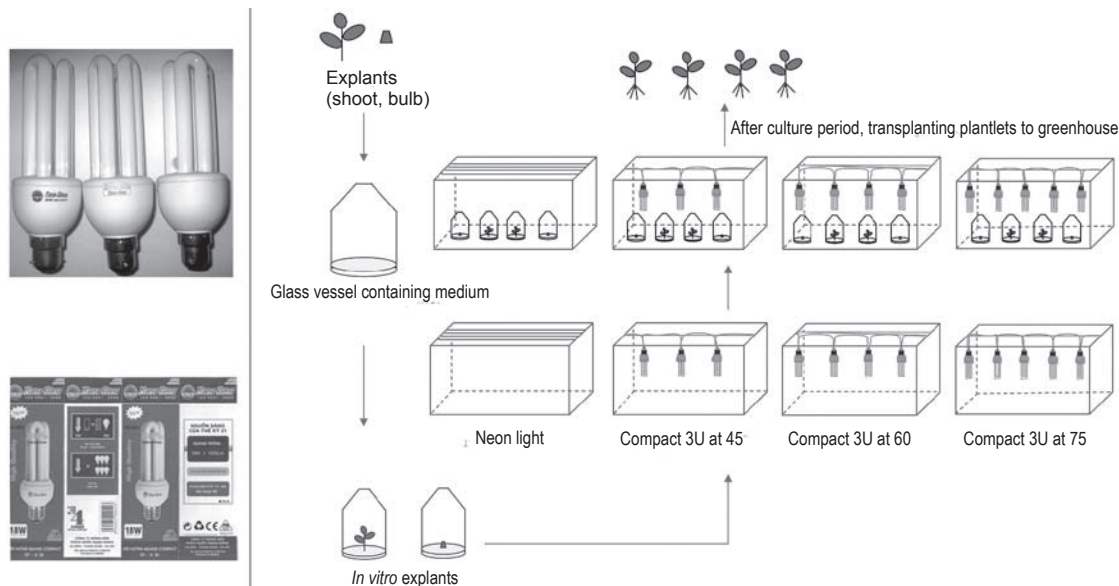


Fig. 1. Compact 3U lamp.

Fig. 2. Setting up the Compact 3U and Neon lighting sources for studying *in vitro* growth and development of some horticultural plants.

height, leaf width, number of leaves, number of roots, root length and plant fresh weight of *Cymbidium* and plant height, bulblet diameter, leaf width, number of leaves, number of roots, root length, and plant fresh weight of *Lilium* were collected.

Culture conditions (*in* and *ex vitro*): *In vitro* cultures were incubated at $25 \pm 2^\circ\text{C}$ with a ten-hour photoperiod and 75-80% relative humidity under different lighting systems as treatment.

After six weeks of culture, *Cymbidium* and *Lilium* plantlets were transplanted to the greenhouse and cultured on tree fern fiber substrate in spongy trays at $25 \pm 2^\circ\text{C}$, 80-85% relative humidity and under 6h/day supplemental Compact 3U lighting source. Plantlets were sprayed with an antifungal solution containing 5 gl^{-1} Dithane M-45 (Dow AgroSciences Co., USA) twice a week. In addition, these plantlets were sprayed with a pesticide solution containing 150 gl^{-1} Sumi alpha (Omo Chemical Ltd., Co., Japan) and fertilizer solution containing 100 gl^{-1} NPK, 50 gl^{-1} Komix BFC 201 (Thien Sinh Biochemical Agriculture and Trade Co., Vietnam) and 15 gl^{-1} Miracle Fort (Phu Hung Foundation, Vietnam) once a week. Moreover, plantlets were also watered twice daily.

Statistical analysis: Each treatment was repeated three times and data was recorded at the 3rd or 6th week of culture. The explants in experiments were arranged in a randomized complete block design with five shoots per treatment and three blocks. The data were analyzed for significance using analysis of variance with the mean separation by Duncan's multiple range test (Duncan, 1995).

Results and discussion

Effect of Compact 3U lighting system on the *in vitro* development of *Fragaria vesca* cv. 'My Da' plantlets: The effect of the Compact 3U and Neon lighting source on the *in vitro* growth and development of strawberry plantlets are described in Table 1 and Fig. 3a. There was a significant difference in plant height, root length, leaf area and plant fresh weight between plantlets placed under two lighting systems at $45 \mu\text{molm}^{-2}\text{s}^{-1}$. The morphological parameters of strawberry cultured under Compact 3U were higher than those under Neon lighting system.

Table 1. Effect of the Compact 3U lighting source on the *in vitro* development of *Fragaria vesca* cv. 'My Da' plantlets after 3 weeks of culture

| Lighting system | Plant height (cm) | Leaf area (mm^2) | Root length (cm) | Plant fresh weight (mg) |
|-----------------|-------------------|-----------------------------|------------------|-------------------------|
| Neon (control) | 3.7b | 45a | 2.5b | 200b |
| Compact 3U | 4.2a | 35b | 3.5a | 300a |

Different letters within a column indicate significant differences ($P = 0.05$) by Duncan's multiple range test.

Table 4. Effect of different irradiances of Compact 3U on the *in vitro* development of *Lilium longiflorum* plantlets after 6 weeks of culture

| Irradiance (Compact 3U) ($\mu\text{molm}^{-2}\text{s}^{-1}$) | Plant height (cm) | Root length (cm) | Leaf area (mm^2) | Number of new formed roots | Number of bulbs | Bulb diameter (mm) | Bulb cluster fresh weight (mg) | Bulb fresh weight (mg) |
|--|-------------------|------------------|-----------------------------|----------------------------|-----------------|--------------------|--------------------------------|------------------------|
| Neon (45) | 9.1c* | 2.5c | 26.1d | 11.1b | 1.9a | 6.4a | 750c | 510c |
| 45 | 11.4a | 3.8a | 40.3a | 10.6c | 1.6c | 5.5d | 810b | 480a |
| 60 | 10.3b | 2.1d | 30.7b | 9.7d | 1.8b | 5.8c | 830b | 470b |
| 75 | 9.1c | 3.0b | 30.0c | 13.1a | 2.0a | 6.1b | 930a | 460b |

* Different letters within a column indicate significant differences ($P = 0.05$) by Duncan's multiple range test

Effect of different irradiances of Compact 3U lighting source on the *in vitro* development of *Cymbidium*, *Lilium* and strawberry plantlets

***Fragaria vesca* cv. 'My Da':** The effect of different Compact 3U irradiances on the *in vitro* development of strawberry is shown in Table 2. In general, *in vitro* strawberry shoots placed under the Neon lighting system had a slower growth than those placed under the Compact 3U lighting system. Two lighting sources had different effects on the morphology and biomass of strawberry, whereas different Compact 3U irradiances virtually did not affect the morphological parameters of strawberry (Fig. 3a). Strawberry plantlets were best at $75 \mu\text{mol m}^{-2}\text{s}^{-1}$, because of the maximum growth and development. However, for commercial purposes, we recommended the use of Compact 3U light at $45 \mu\text{mol m}^{-2}\text{s}^{-1}$ for the micropropagation of strawberry because of saving electrical energy and still remaining the relatively good growth and development.

Table 2. Effect of different irradiances of Compact 3U on the *in vitro* development of *Fragaria vesca* cv. "My Da" plantlets after 3 weeks of culture

| Irradiance (Compact 3U) ($\mu\text{molm}^{-2}\text{s}^{-1}$) | Plant height (cm) | Leaf area (mm^2) | Root length (cm) | Plant fresh weight (mg) |
|--|-------------------|-----------------------------|------------------|-------------------------|
| Neon (45) | 3.7c ^x | 40.5d | 2.5d | 200d |
| 45 | 4.2b | 45.4b | 3.5b | 250c |
| 60 | 4.2b | 41.4c | 3.0c | 270a |
| 75 | 4.4a | 50.0a | 3.9a | 260b |

^x Different letters within a column indicate significant differences ($P = 0.05$) by Duncan's multiple range test.

***Cymbidium* cv. 'Tim Hot':** The effect of different irradiances of Compact 3U on the *in vitro* growth and development of *Cymbidium* plantlets is shown in Table 3. Data showed that there was no significant difference in root length of plantlets placed under the different lighting systems. However, the plantlets were significantly taller at $60 \mu\text{molm}^{-2}\text{s}^{-1}$, with considerably more roots at $45 \mu\text{molm}^{-2}\text{s}^{-1}$ as compared to Neon at $45 \mu\text{molm}^{-2}\text{s}^{-1}$ (Fig. 3b). Besides, the data indicated that the plantlets placed under the Compact 3U lighting system had a better growth than those under the Neon lighting system. The irradiance in this case did not affect the root growth but plantlet fresh weight, which was highest at $60 \mu\text{molm}^{-2}\text{s}^{-1}$. The most suitable irradiance of Compact 3U ($60 \mu\text{molm}^{-2}\text{s}^{-1}$) for growth and morphology of *Cymbidium*, characterized by long shoots and roots, wide leaves and high fresh weight (Fig. 3b), was also the most appropriate intensity

for inducing vigorous growth of the plantlets transplanted to greenhouse.

Table 3. Effect of different irradiances of Compact 3U on the *in vitro* development of *Cymbidium* cv. “Tim Hot” plantlets after 6 weeks of culture

| Irradiance (Compact 3U) ($\mu\text{molm}^{-2}\text{s}^{-1}$) | Plant height (cm) | Root length (cm) | Leaf width (mm) | Number of roots | Plant fresh weight (mg) |
|--|--------------------|------------------|-----------------|-----------------|-------------------------|
| Neon (45) | 10.1b ^w | 3.0a | 4.7b | 2.0c | 620c |
| 45 | 10.2b | 3.0a | 4.8a | 2.5a | 650b |
| 60 | 10.9a | 3.0a | 4.8a | 2.3b | 740a |
| 75 | 10.2b | 2.6b | 4.4c | 2.6a | 610c |

^w Different letters within a column indicate significant differences ($P = 0.05$) by Duncan’s multiple range test.

***Lilium longiflorum*:** The effect of different irradiances of Compact 3U on the *in vitro* growth and development of *L. longiflorum* plantlets is indicated in Table 4 and Fig. 3c. The results suggest that the lighting source as well as its PPF significantly affected the fresh weight of new bulblet clusters derived from initial bulb scales. These results show that *L. longiflorum* bulb scales could be cultured at 75 $\mu\text{molm}^{-2}\text{s}^{-1}$ under Compact 3U for multiplying new high quality bulblets and under Neon at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ for increasing biomass and producing vigorous bulblets before transplanting to greenhouse. In addition, there was a considerable effect of Compact 3U at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ on the *in vitro* *L. longiflorum* morphology. This might be a result of the increase in red light spectrum associated with a low PPF of Compact 3U lighting source which played a certain role in increasing plant height and leaf area. On the other hand, the *L. longiflorum* root morphology was not affected by different lighting sources.

Subsequent growth of *Cymbidium* and *Lilium* plantlets

***Cymbidium* cv. “Tim Hot”:** The subsequent growth of Compact 3U-derived *Cymbidium* plantlets in the greenhouse after one and a half months culture under 6h/day supplemental Compact

Table 5. Subsequent growth of Compact 3U-derived *Cymbidium* plantlets in the greenhouse after one and a half months of culture under 6h/day supplemental Compact 3U lighting source

| Lighting systems ($\mu\text{molm}^{-2}\text{s}^{-1}$) | Plant height (cm) | Leaf width (mm) | Number of leaves | Number of roots | Root length (cm) | Plant fresh weight (g) | |
|---|-------------------|-----------------|------------------|-----------------|------------------|------------------------|------|
| Neon | 45 | 10.4b* | 5.8a | 4.3c | 3.7b | 2.9b | 1.7b |
| | 45 | 10.5b | 5.5b | 4.4c | 3.6b | 3.3a | 1.9a |
| Compact 3U | 60 | 11.1a | 5.4b | 4.9a | 3.7b | 3.2a | 1.8a |
| | 75 | 11.2a | 5.2c | 4.6b | 3.9a | 2.9b | 1.6b |

* Different letters within a column indicate significant differences ($P = 0.05$) by Duncan’s multiple range test.

Table 6. Subsequent growth of Compact 3U-derived *Lilium* plantlets in the greenhouse after 2 months of culture under 6h/day supplemental Compact 3U lighting source

| Lighting systems ($\mu\text{mol m}^{-2}\text{s}^{-1}$) | Plant height (cm) | Bulblet diameter (cm) | Leaf width (mm) | Number of leaves | Number of roots | Root length (cm) | Plant fresh weight (mg) | |
|--|-------------------|-----------------------|-----------------|------------------|-----------------|------------------|-------------------------|------|
| Neon | 45 | 5.5d* | 0.72c | 4.5d | 4.7a | 4.0c | 0.7c | 410c |
| | 45 | 7.6a | 0.77b | 6.3a | 3.5c | 4.0c | 5.6a | 580a |
| Compact 3U | 60 | 6.8b | 0.72c | 6.0b | 3.7b | 4.5b | 1.0b | 560a |
| | 75 | 6.1c | 0.83a | 5.0c | 3.7b | 4.7a | 0.9b | 490b |

* Different letters within a column indicate significant differences ($P = 0.05$) with the mean separation by Duncan’s multiple range test.

3U lighting source at night is given in Table 5. The results showed that *Cymbidium* plantlets grown in the greenhouse under supplemental Compact 3U lighting source had better development than those derived from Neon lighting source (except for leaf width) (Fig. 4a₁, 4a₂).

***Lilium longiflorum*:** Results in Table 6 show that the development of *Lilium* plantlets when transplanted to greenhouse was affected by different lighting sources and intensities. Neon lighting source-derived plantlets had lower growth (plant height and leaf width) but greater number of leaves and better root length than those derived from the Compact 3U lighting source, which yielded variable results under different irradiances (Fig. 4b₁, 4b₂).

In summary, the irradiance of Compact 3U had a positively stimulated impact that affected significantly on the development of these three plants. The Compact 3U lighting source had a positive effect on the plant height, and the plant fresh weight of *Lilium*, *Cymbidium*, the number of strawberry shoots cultured *in vitro* as well as on the root length of strawberry and the number of roots of *Cymbidium* cultured *in vitro*. The results obtained in this study showed that the Compact 3U lighting source affected the morphology of these plants, increased their biomass, and enhanced plantlet growth before transplanting to the greenhouse. Furthermore, this data also showed that different plants adapted differently to different lighting sources, but different irradiances from two lighting sources did not affect root development including root fresh weight, root elongation (except for strawberry) and the number of roots (except for *Cymbidium*) of these plants.

In most cases, different Compact 3U irradiances had no obvious impact on plantlet development as compared to those of Neon light. Different irradiances affected *Lilium* and *Cymbidium* plant height and *Lilium* plant fresh weight. In these plants, a lower intensity gave a higher plant quality. Except for the plant fresh weight of *Lilium* that was enhanced when cultured under Compact 3U at 75 $\mu\text{molm}^{-2}\text{s}^{-1}$, the remaining cases showed that plantlets developed well under lower intensities (45 or 60 $\mu\text{mol m}^{-2}\text{s}^{-1}$). Consequently, Compact 3U confirmed the positive effect

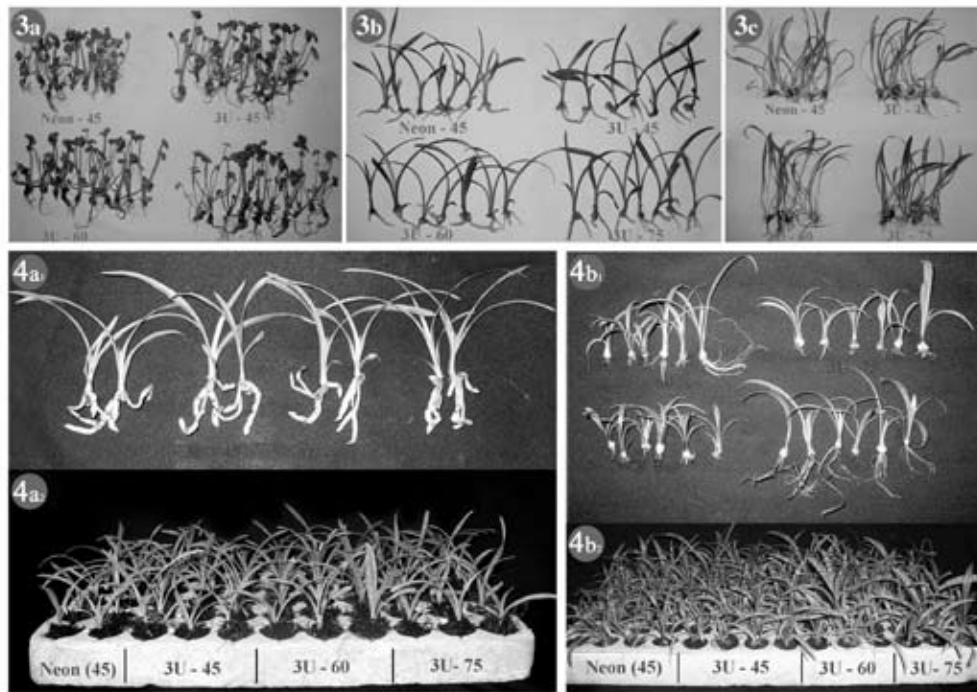


Fig. 3. Strawberry, *Cymbidium* and *Lilium* plantlets cultured under Compact 3U at different irradiances. (a): Strawberry cultured under Neon light at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ (top left), Compact 3U at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ (top right), 60 $\mu\text{molm}^{-2}\text{s}^{-1}$ (bottom left), or 75 $\mu\text{molm}^{-2}\text{s}^{-1}$ (bottom right) after three weeks of culture. (b): *Cymbidium* plantlets cultured under Neon light at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ (top left), Compact 3U at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ (top right), 60 $\mu\text{molm}^{-2}\text{s}^{-1}$ (bottom left), or 75 $\mu\text{molm}^{-2}\text{s}^{-1}$ (bottom right) after six weeks of culture. (c): *Lilium* plantlets cultured under Neon light at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ (top left), Compact 3U at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ (top right), 60 $\mu\text{molm}^{-2}\text{s}^{-1}$ (bottom left), or 75 $\mu\text{molm}^{-2}\text{s}^{-1}$ (bottom right) after six weeks of culture.

Fig. 4. *Cymbidium*, *Lilium* plants after transplanted in greenhouse. (a.): *Cymbidium* plants transplanted in greenhouse after one and a half months. Left to right: *Cymbidium* plants derived from Neon light at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$, Compact 3U at 45, 60 or 75 $\mu\text{molm}^{-2}\text{s}^{-1}$. (a.): *Cymbidium* plants in spongy tray after one and a half months transplanted in greenhouse. Left to right: *Cymbidium* plants derived from Neon light at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$, Compact 3U at 45, 60 or 75 $\mu\text{molm}^{-2}\text{s}^{-1}$. (b.): *Lilium* plants derived from Neon light at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ (bottom left), Compact 3U at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$ (bottom right), 60 $\mu\text{molm}^{-2}\text{s}^{-1}$ (top left), or 75 $\mu\text{molm}^{-2}\text{s}^{-1}$ (top right) after two months in greenhouse. (b.): *Lilium* plants in spongy tray after two months transplanted in greenhouse. Left to right: *Lilium* plants derived from Neon light at 45 $\mu\text{molm}^{-2}\text{s}^{-1}$, Compact 3U at 45, 60 or 75 $\mu\text{molm}^{-2}\text{s}^{-1}$.

on the *in vitro* development of plantlets before transplanting to greenhouse.

These above results were similar to those of Warrington and Michell (1976), Morgan and Smith (1981), Smith (1982), Tibbitts *et al.* (1983), Mortensen and Stromme (1987), Economou and Read (1987), and Agrawal (1992), who all confirmed the significant effects of light quality (related to different lighting sources) on the growth and morphology of *in* and *ex vitro* plants. The effect of irradiance of different lighting sources on the development of plants were also the concern of some studies of Gilslerod and Mortensen (1997), Miyashita *et al.* (1997), and Nhut (2002). In these studies, higher irradiances gave the best plant growth. But in this report, we suggest for use of lower intensities (45 or 60 $\mu\text{molm}^{-2}\text{s}^{-1}$) of Compact 3U for plant propagation owing to the reduction of electrical energy consumption as well as the increase in the development of some horticultural plants.

The Compact 3U lighting source had a highly significant effect on the development of *Cymbidium* 'Tim Hot', *Lilium longiflorum* and *Fragaria vesca* cv. 'My Da' as in this study. The results indicate that these plants adapted differently with different light sources and intensities. Strawberry shoots had better growth and morphology when cultured under Compact 3U lighting system than those under Neon light system. The preeminence of Compact 3U lighting system was also expressed when shoots of three plants were cultured under different irradiances of two lighting system.

Though in most cases, different irradiances of Compact 3U had no obvious effect on the development of plantlets as compared to the Neon light, lower irradiance gave higher *Cymbidium* and *Lilium* plantlets quality. Moreover, plantlets derived from Compact 3U lighting system developed better than those from Neon lighting system.

Hence, the Compact 3U lamp, having a suitable light spectrum, resulting in good, high quality plants, saving 75-80% of electrical energy consumption as compared to incandescent lamps, being cheap, and subsequently retrieving initial investments quickly, was expected to be a novel lighting system used in the successful micropropagation and subsequent *ex vitro* growth of some horticultural plants. In fact, because of obvious advantages, the Compact 3U lamp has been used as the lighting source for propagation of many horticulture plants in Dalat, Lam Dong, Vietnam.

References

- Agrawal, S.B. 1992. Effects of supplemental UV-B radiation on photosynthetic pigment, protein and glutathione contents in green algae. *Env. Exp. Bot.*, 32(2): 137-143.
- Collin, R.N., E.N. Margaret, H. Ted and P. David, 1988. Light quality and light pipes in the micropropagation of woody ornamental plants. *Acta Hortic.*, 226: 413-416.
- Debergh, P.C. and R. Zimmerman, 1990. Micropropagation: technology and application. *Kluwer Academic Publishers, Dordrecht*, pp. 484.

- Duncan, D.B. 1995. Multiple range and multiple F test. *Biomet.*, 11: 1-42.
- Economou, A.S. and P.E. Read, 1987. Light treatments to improve efficiency of *in vitro* propagation systems. *HortSci.*, 22(5): 751-754.
- Faust, J.E. and R.D. Heins, 1997. Quantifying the influence of high-pressure sodium lighting on shoot-tip temperature. *Acta Hort.*, 418: 85-92.
- Gabarkiewicz, B., E. Gabryszewska, R. Rudnicki and D. Goszcynska, 1997. Effects of light quality on *in vitro* growing of *Dieffenbachia* cv. compacta. *Acta Hort.*, 418: 159-162.
- Gabryszewska, E. and R.M. Rudnicki, 1997. The effects of light quality on the growth and development of shoots and roots of *Ficus benjamina* *in vitro*. *Acta Hort.*, 418: 163-168.
- Gilslerod, H.R. and L.M. Mortensen, 1997. Effect of light intensity on growth and quality of cut roses. *Acta Hort.*, 418: 25-32.
- Hart, J.W. 1988. Light and Plant Growth. In: *Topics in Plant Physiology*, 1: 34-50.
- Hayashi, M., N. Fujita, Y. Kitaya and T. Kozai, 1992. Effect of sideward lighting on the growth of potato plantlets *in vitro*. *Acta Hort.*, 319: 163-166.
- Iwanami, Y., T. Kozai, Y. Kitaya and S. Kino, 1992. Effects of supplemental red and far-red lighting using light emitting diodes on stem elongation and growth of potato plantlets *in vitro* (Abstr.). *International Symposium Transplant production Systems, Yokohama, Japan*, pp. 183.
- Kim, H.H. and T. Kozai, 2000. Production of value-added transplants in closed systems with artificial lighting. In: *Transplant Production in the 21st Century*. C. Kubota and C. Chun (eds.), Kluwer Academic Publishers, 137-144.
- Kirdmanee, C., Y. Kitaya and T. Kozai, 1993. Effect of supplemental far-red lighting and photosynthetic photon flux density on stem elongation and dry weight increase of *Eucalyptus camaldulensis* *in vitro* plantlets (Abstr.). XVth International Botanical Congress Yokohama, Japan, pp. 537.
- Kozai, T., K. Fujiwara, M. Hayashi and J. Aitken-Christie, 1992. The *in vitro* environment and its control in micropropagation. *Transplant production systems*. Kluwer Academic Publishers. Dordrech, pp. 247-282.
- Kunneman, B.P.A.M. and J.B. Ruesink, 1997. Interactions between light, temperature and CO₂ in rooting of conifer cuttings. *Acta Hort.*, 418: 97-102.
- Maas, F.M. and E.J. Bakx, 1997. Growth and flower development in roses as affected by light. *Acta Hort.*, 418: 127-134.
- Miyashita, Y. 1995. Growth and morphology of potato plantlets *in vitro* as affected by light quality, and size and nodal position of the explant. *Graduate School of Science and Technology*. Chiba University, pp. 1-5.
- Miyashita, Y., T. Kimura, Y. Kitaya, C. Kubota and T. Kozai, 1997. Effects of red light on the growth and morphology of potato plantlets *in vitro*: Using light emitting diodes (LEDs) as a lighting source for micropropagation. *Acta Hort.*, 418: 169-176.
- Moe, R. 1997. Physiological aspects of supplementary lighting in horticulture. *Acta Hort.*, 418: 17-24.
- Morgan, D.C. and H. Smith, 1981. Non-photosynthetic responses to light quality. In: *Encyclopedia of Plant Physiology*. Springer-Verlag, Berlin, pp. 109-134.
- Mortensen, L. M. and E. Stromme, 1987. Effects of light quality on some greenhouse crops. *Sci. Hort.*, 33: 27-36.
- Murakami, K., I. Aiga, K. Horaguchi and M. Morita, 1997. Red/far-red photon flux ratio used as an index number for morphological control of plant growth under artificial lighting conditions. *Acta Hort.*, 418: 135-140.
- Murashige, T. and F. Skoog, 1962. A revised medium for rapid growth and bioassays with tobacco tissue culture. *Plant Physiol.*, 15: 473-497.
- Nhut, D.T. 2002. *In vitro* growth and physiological aspects of some horticultural plantlets cultured under red and blue light emitting diodes (LEDs). PhD Thesis, Kagawa University, pp. 163-172.
- Schwabe, W.W. 1963. Morphogenetic responses to climate. In: *Environmental Control of Plant Growth*. Academic Press, New York, pp. 311-336.
- Seabrook, J.E.A. 1987. Changing the growth and morphology of potato plantlets *in vitro* by varying the illumination source. *Acta Hort.*, 212: 401-410.
- Smith, H. 1982. Light quality, photoperception and plant strategy. *Annual Rev. Plant Physiol.*, 33: 481-518.
- Tibbitts, T.W., D.C. Morgan and I.J. Warrington, 1983. Growth of lettuce, spinach, mustard, and wheat plants under four combinations of high-pressure sodium, metal halide, and tungsten halogen lamps at equal PPFD. *J. Amer. Soc. Hort. Sci.*, 108(4): 622-630.
- Walz, F. and W. Horn, 1997. The influence of light quality on gas exchange of dendranthema. *Acta Hort.*, 418: 53-58.
- Warrington, I.J. and K.J. Michell, 1976. The influence of blue- and red-biased light spectra on the growth and development of plants. *Agr. Met.*, 16: 247-262.
- Wulster, G. and H.W. Janes, 1997. Effects of supplemental light quality, quantity, and differential temperature on growth and development of Easter Liliu (*Lilium longiflorum*). *Acta Hort.*, 418: 153-158.