

# Relative uptake of the fungicide carbendazim by selected fruits and vegetables and keeping quality of apple and tomato after dip treatment

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# Abstracts

Uptake of fungicide carbendazim by eight fruits and vegetables on dipping in carbendazim aqueous suspension, and its effects on keeping quality of tomato and apple were investigated. The uptake of carbendazim varied significantly (p<0.01), ranging from 68.97±2.89 to 813.64±11.46 µg (mean 342.13 µg) among the fruits and vegetables. The lowest uptake was in apple followed by banana, orange, tomato, okra, grape (golden), grape (blue), sapota and carrot. The dip treatment was more effective for storage life extension of tomato than apple and at ambient ( $32\pm2^{\circ}$ C) than at low temperature ( $7\pm2^{\circ}$ C). Cumulative physiological loss in weight, physical appearance and spoilage in tomato and apple, and lycopene content, titratable acidity, ascorbic acid content and moisture content in tomato were also analysed during the course of storage.

Key words: Carbendazim uptake, fruits and vegetables, dip treatment, keeping quality

# Introduction

Benomyl (IUPAC: Methyl 1 (Butyl carbamoyl) benzimidazole 2 yl carbamate) and carbendazim or MBC (IUPAC: Methyl benzimidazole 2 yl carbamate), two important benzimidazole class of fungicides, are applied predominantly on fruits and vegetables in pre- as well as post-harvest stages (Ben-Yeshoshua and Cohen, 1981; Mohammed and Sealy, 1988; Gupta and Mukherjee, 1980; Awasthi and Sharma, 1997). Benomyl and other benzimidazole fungicides, as such or after hydrolysis to MBC inside the plant body, leave stable residues, having long term toxicity to growing cells and inhibitory effect on DNA, RNA and protein synthesis by inhibiting cell division (Charles, 1987).

Monitoring of farm gate samples in India, of late, has shown carry-over pesticide residues to an extent of 40% in fruits (Agnihotri, 1999) and 60% in vegetables (Ahuja et al., 1998; Awasthi and Ahuja, 1997), showing contamination from preharvest application of pesticides including benomyl and MBC. On the other hand, benomyl and MBC are applied post-harvest to control fungal decay in a number of fruits and vegetables, and hence to extend their shelf-life (Waskar et al., 1999a, 1999b; Amadioha, 1996, 1998; Sanderson, 1999; Nwufo et al., 1994; Jahangir et al., 1994; Abu-Baker and Abdul-Karim, 1994; Papadopoulou-Mourkidou, 1991; Goulart et al., 1992; Rathu et al., 1989; Sharma et al., 1989). Due to high persistence of carbendazim, its residues have been reported in many fruits and vegetables (USDA, 1993; Awasthi 1998; Awasthi and Ahuja, 1997; Awasthi and Sharma, 1997; Baldi et al., 1981, 1982; Bencivenga and et al., 1982; Cano and Plaza, 1987; Hamil and Harper, 1982; Hargreaves, 1983; Kiigemagi et al., 1991; Meloni and Pirisi, 1984; Monico-Pifarrae, 1987; Nagayama et al., 1983).

But there is no systematic study carried out on uptake of MBC residues due to dip treatment of fruits and vegetables. In the present study, an attempt has been made to study the extent of uptake of MBC residue by eight common fruits and vegetables

dipped in MBC suspension under laboratory conditions. Effect of dip treatment using MBC suspension, on extension of storage life of tomato and apple was investigated.

# Materials and methods

Selection of fruits and vegetables: Eight common fruits and vegetables *e.g.* tomato (*Lycopersicon esculentum* L.), apple (*Malus pumila*), carrot (*Daucus carrota* L.), okra (*Abelmoschus esculentus* Moench.), orange (*Citrus sinensis* Osbeck.), grapes (*Vitis vinifera* L.) of golden yellow and blue colour, sapota (*Achras zapota* L.) and banana (*Musa paradisiaca* L.), which predominantly get the application of benomyl or MBC in preharvest and occasionally in post-harvest stages, were selected for the present study. Fresh, fully mature firm medium ripe tomato (S-15 variety), freshly harvested carrot and tender okra, fully mature and hard-ripe sapota and fully mature medium ripe banana obtained as farmgate samples were selected, whereas firm and full apple (Golden Delicious), ripe orange and fully mature ripe grapes purchased from local market were used.

**Dip treatment**: Whole fruit or vegetable without any structural damage or injury was weighed (~1500 g) and immersed completely for 1 hr at ambient temperature in 2000 ml aqueous suspension of 750 ppm Bavistin 50% WP. (*i.e.*, 375 ppm of carbendazim as *a.i.*), taken in a stainless steel vessel covered with a lid. After 1 hr, the fruit or the vegetable was freed of the suspension and air-dried in laboratory condition till there was no trace of visible water on the surface. The weight of the fruit or the vegetable and the volume of the suspension before and after dip treatment were recorded to calculate percent gain in weight by the treated fruits or the vegetables and loss in volume of MBC suspension.

**Analysis of fruits and vegetables for MBC:** The fruits or the vegetables prior and after dip treatment were analysed for MBC residue by modifying the method of Rangaswamy *et al.* (1987). The difference in the carbendazim residue contents was

considered as the residue uptake, and expressed as  $\mu g$  per 100 g sample,  $\mu g$  per 100 g MBC and  $\mu g$  per 100 ml MBC suspension.

**Studies on storage life:** The spiked and the non-spiked tomato and apple samples were stored at  $32\pm2^{\circ}$ C (ambient) and  $7\pm2^{\circ}$ C (low) to observe cumulative physiological loss in weight (CPLW), skin shrivelling and fungal spoilage. The period in days just prior to appearance of fungal spoilage was considered as the storage life. Percent spoilage of tomato and apples was calculated by expressing the ratio of the weight of the spoiled tomato or apple to the weight of tomato or apple taken for storage study in percentage. The CPLW was determined by the difference in weight between the fruits or vegetables of the first day and that of the any respective storage day to get the CPLW of the particular day.

**Chemical quality analysis:** Chemical quality parameters such as moisture content, titratable acidity, ascorbic acid and lycopene contents of the fresh as well as the stored samples of the treated and the control tomatoes were analysed by the standard oven dry (AOAC, 1990), titrimetric (AOAC, 1990) and spectrophotometric (Beerh and Siddappa, 1959) methods, respectively.

**Statistical analysis:** All the data were subjected to calculation of mean±standard deviation of a number replicates and subsequent one-way analysis of variance (ANOVA) for significant differences among them (Snedecor and Cochran, 1989)

### **Results and discussion**

**Uptake of MBC residue:** Mean weights of fruits and vegetables samples before and after dip treatment ranged from  $1500.5\pm0.2$  to  $1519.6\pm2.4$  g (mean 1508.67g) and  $1509.1\pm2.1$  to  $1536.1\pm2.5$ g (1523.38g), respectively (Table 1). Gain in weight by the samples due to dip treatment varied from  $0.41\pm0.02$  to  $1.55\pm0.09\%$  (mean 0.97%) depending upon the type of fruits and vegetables (Table 1). On the other hand, mean volumes of MBC suspension before and after dip treatment were  $2000\pm0.00$  ml (mean 2000 ml) and  $1942.7\pm2.6$  to  $1979.2\pm1.2$  ml (mean 1962.82 ml), respectively (Table 1). Decrease in volume of MBC suspension used for dip treatment ranged from  $1.04\pm0.05$  to  $2.86\pm0.15\%$  (mean 1.86%) depending upon the type of fruits and vegetables (Table 1).

Percentage weight gain during dip treatment was considered as actual uptake of MBC suspension by the fruits and vegetables, whereas percentage decrease in volume of MBC suspension included the loss due to actual uptake of MBC suspension as well as the loss due to evaporation or spillage, etc. at the time of handling the produce. Hence, the percentage weight gain represented the uptake of MBC residue more appropriately than the percentage volume loss. The percentage weight gain values were 39.42 to 72.95% (mean 51.87%) lower than the percentage loss in volume. The fruits and vegetables like tomato, grapes, apple with waxy smooth skin showed less gain in weight than the ones like carrot, sapota, banana, okra, etc. with rough skin. Because uptake of residues was due to penetration of residue through skin or peel.

Quantity of MBC uptake (ppm) was found to vary significantly (p<0.01) among the fruits and vegetables as the case may be, and ranged from  $68.97\pm2.89$  to  $813.64\pm11.46$  µg (mean 342.13 µg) (Table 1). Uptake of MBC residue expressed as µg per100g sample, µg per100 µg MBC and µg per100 ml suspension was also found to vary significantly (*p*<0.01) from one fruit or vegetable to others (Table 1). However, uptake of MBC residue (expressed in all three

different forms) was the lowest in apple followed by (in an increasing order), banana, orange, tomato, okra, grape (golden), grape (blue), sapota and carrot. Lower uptake in grape was due to poor penetration of suspension through peels. During spiking the residue might have deposited on the surface, leading to slight absorption by outer waxy layers and subsequently inner cuticles. A similar observation was made on absorption of field-applied pesticides on standing crops (Cabras *et al.*, 1998). In a detailed study on mechanism of pesticide absorption on plant body, enough evidence was presented to show that the driving force for pesticide penetration into plant body depended on formulation, lipophilicity and concentration of the active ingredient (Baur *et al.*, 1997; Marzouk *et al.*, 1998).

**Physical change or fungal spoilage:** Shrivelling of skin started on 5<sup>th</sup> and 10<sup>th</sup> day in case of the spiked tomato and 4<sup>th</sup> and 8<sup>th</sup> day in case of the non-spiked one at AT and LT, respectively (Table 2). On the other hand, initiation of fungal spoilage was observed on 9<sup>th</sup> and 18<sup>th</sup> day for the spiked tomato and 6<sup>th</sup> and 14<sup>th</sup> day for the non-spiked one at AT and LT, respectively (Table 2). It showed that initiation of fungal spoilage in tomato was delayed by the MBC treatment to a greater extent at LT than that at AT. Moreover, percentage spoilage was found to be significantly (p<0.01) less in the spiked tomato than the non-spiked one on 10<sup>th</sup> and 22<sup>nd</sup> day of storage at AT and LT, respectively (Table 2).

Storage life: Although skin shrivelling set in earlier than initiation of fungal decay, storage life of tomatoes and apples were determined by initiation of fungal spoilage and the appearance of black spot on skin as an indication, respectively. Both the spiked and the non-spiked apples showed significantly (p < 0.01) longer storage life than the spiked and the non-spiked tomato, respectively at AT as well as LT (Table 2). Furthermore, the spiked tomato were found to have a storage life of 9 and 18 days at RT and LT respectively, which were significantly (p < 0.01) greater than the respective storage life of the non-spiked tomato (Table 2). It showed that dip treatment with MBC extended the storage life of tomato significantly (p < 0.01) at both the storage temperatures. Use of benomyl or MBC as a post-harvest treatment for extension of storage life of a number of fruits and vegetables was reported earlier (Abu-Baker and Abdul-Karim, 1994; Amadioha, 1996, 1998; Goulart et al., 1992; Jahangir et al., 1994; Nwufo et al., 1994; Papadopoulou-Mourkidou, 1991; Rathu et al., 1989; Sanderson, 1999; Sharma et al., 1989; Waskar et al., 1999a, 1999b).

**Cumulative physiological loss in weight (CPLW):** Percentage CPLW was found to be significantly (p < 0.01) greater in tomatoes than in apples at both the storage temperatures. Values of CPLW were more at AT than at LT for both the spiked and the non-spiked tomatoes and apples on the same days of storage (Table 2). Physiological loss in weight in fruits and vegetables is due to respiration and evapo-transpiration, which in turn depends on atmospheric temperature and humidity and number of lenticels or stomata per unit area of the fruits and vegetables. Furthermore, the treated samples of both tomatoes and apples showed significantly (p < 0.01) lower % CPLW than the corresponding non-treated sample during the course of storage at both the temperatures (Table 2).

**Moisture content (wet weight basis):** Percent moisture content of the treated and the control tomatoes was  $95.03\pm0.49$ , which decreased significantly (p < 0.01) after 4 and 8 days of storage at

Table 1.	Uptake o	f MBC resid	ue by differ	ent fruits and	I vegetables									
Samples	5	Veight of Sa	mple (g)		Volume	s of MBC su	spension	Conc	centration	Amount	Upta	ke of MBC by variou	S	Average
	Before	Aft	эr	Gain in	-	(375 ppm) r	lu	0	f MBC	of MBC	μ	its and vegetables		recovery
	spiking	spiki	ing	weight	Before	After	Gain	ر يا	ıptake	used	µg/100	μg/100	μg/100 €	xtraction
				(%)	spiking	spiking	weig	ght	(ppm)	шg	sample	µg MBC	ml MBC	(%)
Tomato	1503.1+1	6 1509.1	+2.1	0.41+0.02 <sup>a</sup>	2000+0.00	1979.2+1	2 1.04+	0.05 <sup>a</sup> 3	75+0.00	276.55+4.65 <sup>d</sup>	18.40+0.31	1 0.0369+0.0006 <sup>d</sup>	13.82+0.23 <sup>d</sup>	
Apple	1519.6±2.	4 1531.7	+2.9	$0.79\pm0.03^{cd}$	$2000\pm0.00$	1961.4±1.	9 1.93±	0.09 <sup>d</sup> 3	75±0.00	68.97±2.89 <sup>a</sup>	$4.54\pm0.19$	a 0.0092±0.0004 <sup>a</sup>	$3.45\pm0.14^{a}$	
Carrot	1507.8±0.	8 1531.2	±1.3	$1.55\pm0.09^{f}$	2000±0.00	1946.2±2.	3 2.69±	0.12 <sup>e</sup> 3	75±0.00 8	13.64±11.46 <sup>i</sup>	$53.96\pm0.76$	$0.1085\pm0.0015^{\circ}$	40.68±0.57 <sup>i</sup>	
Okra	1500.8±0.	5 1518.6	<b>主1.0</b>	1.19±0.11 <sup>e</sup>	2000±0.00	1962.6±1.	7 1.87±	0.09 <sup>d</sup> 3	75±0.00	303.44±4.03 <sup>e</sup>	20.22±0.27	0.0405±0.0005 <sup>e</sup>	15.19±0.20 <sup>e</sup>	
Orange	1514.2±2.	2 1528.0	)±2.7	0.91±0.10°	2000±0.00	1960.3±3.	0 1.98±	0.13 <sup>d</sup> 3	75±0.00	189.41±3.44 <sup>c</sup>	$12.51\pm0.23$	° 0.0253±0.0005°	9.47±0.17 <sup>c</sup>	79.39
Grapes <sup>&amp;</sup>	1501.6±0.	2 1510.8	J±0.6	0.61±0.04 <sup>b</sup>	2000±0.00	1974.4±1.	5 1.28±	0.08 <sup>b</sup> 3	75±0.00	328.63±3.09 <sup>f</sup>	21.89±0.21	$0.0438\pm0.0004^{f}$	16.43±0.15 <sup>f</sup>	
Grapes <sup>&amp;&amp;</sup>	1500.5±0.	2 1511.2	±0.5	0.71±0.04 <sup>c</sup>	2000±0.00	1970.5±1.	4 1.47±	0.09 <sup>bc</sup> 3	75±0.00	342.55±3.41 <sup>g</sup>	22.83±0.23	<sup>3</sup> 0.0457±0.0005 <sup>9</sup>	17.13±0.17 <sup>9</sup>	
Sapota	1511.9±1.	5 1533.7	'±1.9	1.44±0.09 <sup>f</sup>	2000±0.00	1942.7±2.	6 2.86±	0.15 <sup>e</sup> 3	75±0.00 (	571.93±9.91 <sup>h</sup>	44.44±0.66	0.0896±0.0013 <sup>h</sup>	33.60±0.50 <sup>h</sup>	
Banana	1518.5±2.	4 1536.1	±2.5	1.16±0.06 <sup>e</sup>	2000±0.00	1968.1±1.	1 1.59±	0.06 <sup>c</sup> 3	75±0.00	84.06±3.12 <sup>b</sup>	5.54±0.21	<sup>o</sup> 0.0112±0.0004 <sup>b</sup>	4.20±0.16 <sup>b</sup>	
Mean	1508.67	1523.	38	0.97	2000	1962.82	1.8(	9	375	342.13	20.70	0.0456	17.11	
Mean ± St	andard De	viation values	s with differer	nt superscripts	a, b, c, diff	er significant	ly ( <i>p</i> <0.01). &≓l	pale to green	ı, &&=blue bla	ack				
Table 2. E	Effect of <b>N</b>	MBC treatm€	ent, storage	period and to	emperature o	m CPLW(%)	), physical ch	ange or spo	oilage and s	torage life of t	omato and a	pple		
Sample	Storage	0		Percer	nt CPLW on s	storage at d	ifferent tempe	erature on o	different day	S		Physical change		Storage
	temp*	2	4	9	8	12	16	20	24	28	32	and fungal spoil	age I	ife(Day)**
Spiked	АТ	3.28±0.07 <sup>c</sup>	5.76±0.03 <sup>1</sup>	8.59±0.07 <sup>k</sup>	10.95±0.14 <sup>q</sup>	QN	ΠN	QN	ND	DN	S ON	kin shrivelling started	d on 5th day;	ЭB
tomato											52	oulage started on sun d reight basis) spoiled ol	iay ariu oo.o 7⁄o n 10 <sup>th</sup> day.	
	Ц	1.33±0.00 <sup>a</sup>	2.31±0.06 <sup>g</sup>	3.14±0.03°	4.02±0.04p	5.04±0.05 <sup>t</sup>	5.87±0.02 <sup>u</sup> (	6.41±0.04 <sup>j</sup>	QN	QN	ND DN	kin shrivelling started ngal spoilage started or (weight basis) spoiled	on 10th day; n 18th day; 8.8 1 on 22 <sup>nd</sup> day	18 <sup>E</sup>
Non-spikec tomato	ł AT	3.84±0.02 <sup>d</sup>	6.69±0.06 <sup>j</sup>	9.31±0.06 <sup> </sup>	11.78±0.19 <sup>r</sup>	Ŋ	QN	QN	QN	QN		kin shrivelling started on boilage started on 6 <sup>th</sup>	dth day; funga day; 82.1 % 10th day	6A
	Ц	1.98±0.05 <sup>b</sup>	2.81±0.03 <sup>i</sup>	3.71±0.07 <sup>d</sup>	4.63±0.06°	5.99±0.04 <sup>u</sup>	6.79±0.04 <sup>i</sup> 7	′.31±0.08 <sup>y</sup>	QN	QN	DN DN DN	kin shrivelling started ngal spoilage started or (weight basis) spoiled	1 on 8th day; 14 <sup>th</sup> day; 89.6 d on 22 <sup>nd</sup> day.	14 <sup>C</sup>
Spiked app	ile AT	QN	1.98±0.01 <sup>b</sup>	QN	3.76 ±0.05 <sup>d</sup> .	4.47±0.07°	6.02±0.16 <sup>u</sup> 8	3.30±0.11 <sup>k</sup>	QN	QN		kin shrivelling started ack spot appeared on	on 10th day; 16 <sup>th</sup> day.	16 <sup>D</sup>
	Ц	Ŋ	0.69±0.00 <sup>e</sup>	QN	1.34±0.02 <sup>a</sup>	1.61±0.02 <sup>s</sup>	2.41±0.03 <sup>∨</sup> 2	2.78±0.01 <sup>i</sup>	3.10±0.02 <sup>c</sup>	3.37±0.05° 3	.72±0.05 <sup>d</sup> S bl	kin shrivelling started or ack spot appeared on	1 30th day; 35 <sup>th</sup> day.	35 <sup>G</sup>
Non-spikec apple	I AT	QN	2.13±0.04 <sup>9</sup>	QN	4.17±0.02 <sup>n</sup>	5.11±0.00 <sup>t</sup>	7.86±0.06 <sup>x</sup> 9	9.37±0.13 <sup> </sup>	QN	QN	ND ND	kin shrivelling started or ack spot appeared on	n 10th day; 14 <sup>th</sup> day.	14 <sup>C</sup>
	Ц	QN	0.78±0.02 <sup>f</sup>	QN	1.48±0.00 <sup>m</sup>	1.98±0.04 <sup>b</sup>	2.55±0.05 <sup>w</sup> 2	2.94±0.01 <sup>z</sup>	3.24±0.04 <sup>c</sup>	3.68±0.04 <sup>d</sup> 4	.03±0.02 <sup>p</sup> S bl	kin shrivelling started or ack spot appeared on	n 30th day; 32nd day.	$32^{F}$
AT - Ambie **Mean val	ent temper ues with d	ature (32±2°⊧ ïfferent super	C), LT - Low scripts A, B,	temperature ( C,vary sign	7±2°C), ND - ìificantly (p<0.(	·Not done*M 01).	ean ± Standarc	d Deviation v	alues with dif	ferent superscri	pts a, b, c,	differ significantly (p<	0.01).	

	Table 3.	Effect of MBC treatmen	t, storage	period and tem	perature on p	percent moisture o	content and acidit	ty of tomato
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Samples Storage Storage period (days)							
·	Temperature	0	4	8	12	16	24
Percent moist	ure content (	wet weight basis					
Spiked tomato	AT	95.03±0.49 <sup>Aa</sup>	92.06±0.26 <sup>Bbc</sup>	91.14±0.31 <sup>CFc</sup>	91.02±0.20 <sup>Cc</sup>	ND	ND
	LT	95.03±0.49 <sup>Aa</sup>	93.65±0.40 <sup>Dabc</sup>	92.77±0.43 <sup>Deb</sup>	92.41±0.23 <sup>EHb</sup>	92.19±0.27 <sup>EHb</sup>	92.02±0.31 <sup>EHbc</sup>
Non-spiked toma	ato AT	95.03±0.49 <sup>Aa</sup>	91.88±0.29 <sup>EFbc</sup>	90.79±0.28 <sup>Gc</sup>	90.63±0.34 <sup>Gc</sup>	ND	ND
·	LT	95.03±0.49 <sup>Aa</sup>	93.42±0.38 <sup>Dab</sup>	92.69±0.24 <sup>DEab</sup>	92.26±0.19 <sup>EHb</sup>	91.94±0.27 <sup>FHbc</sup>	91.78±0.22 <sup>Hbc</sup>
Percent titrata	ble acidity as	citric acid (dry v	weight basis)				
Spiked tomato	AT	7.63±0.24 <sup>Åa</sup>	9.38±0.32 <sup>Bbc</sup>	10.07±0.42 <sup>BCbc</sup>	10.46±0.36 <sup>Cc</sup>	ND	ND
	LT	7.63±0.24 <sup>Aa</sup>	7.88±0.19 <sup>Aa</sup>	8.19±0.17 <sup>ADFa</sup>	8.59±0.21 <sup>DEFa</sup>	8.91±0.29 <sup>BEFb</sup>	9.45±0.25 <sup>BFbc</sup>
Non-spiked toma	ato AT	7.63±0.24 <sup>Aa</sup>	9.56±0.24 <sup>BCbc</sup>	10.43±0.30 <sup>Cc</sup>	10.66±0.44 <sup>Cc</sup>	ND	ND
	LT	7.63±0.24 <sup>Aa</sup>	7.96±0.26 <sup>ADa</sup>	8.28±0.18 <sup>DFa</sup>	8.71±0.33 <sup>BFGa</sup>	9.06±0.20 <sup>FBb</sup>	9.52±0.32 <sup>BGbc</sup>

Table 4. Effect of MBC treatment, storage period and temperature on lycopene and ascorbic acid contents of tomato

Samples	Storage		St	orage period (da	eriod (days)			
	temp	0	4	8	12	16	24	
Lycopene conter	nt (mg pei	r 100 g dry weight	)					
Spiked tomato	AT	34.41±3.22 <sup>Aa</sup>	85.77±4.66 <sup>BHcd</sup>	106.79±3.51 <sup>Cef</sup>	119.60±3.56 <sup>DFfghi</sup>	ND	ND	
	LT	34.41±3.22 <sup>Aa</sup>	44.88±3.31 <sup>Eab</sup>	79.81±2.07 <sup>Bc</sup>	105.93±3.95 <sup>Cef</sup>	116.90±2.69 <sup>Dfgh</sup>	127.19±3.51 <sup>Fghi</sup>	
Non-spiked tomato	AT	34.41±3.22 <sup>Aa</sup>	108.50±4.68 <sup>Cdefg</sup>	117.92±2.50 <sup>Dfgh</sup>	131.59±4.59 <sup>Gh</sup>	ND	ND	
	LT	34.41±3.22 <sup>Aa</sup>	49.10±3.19 <sup>Ea</sup>	94.12±2.60 <sup>Hde</sup>	126.49±3.36 <sup>FGgh</sup>	129.53±2.55 <sup>FGhi</sup>	132.24±3.60 <sup>Fgi</sup>	
Ascorbic acid co	ontent (m	g per 100 g dry we	eight)					
Spiked tomato	AT	423.46±6.31 <sup>Åa</sup>	264.11±4.75 <sup>Bc</sup>	188.21±3.29 <sup>Cf</sup>	151.09±2.87 <sup>Dg</sup>	ND	ND	
	LT	423.46±6.31 <sup>Aa</sup>	379.30±4.84 <sup>Eb</sup>	291.56±3.64 <sup>Fe</sup>	262.19±3.95 <sup>Bc</sup>	218.51±2.44 <sup>Gh</sup>	172.17±2.26 <sup>Hi</sup>	
Non-spiked tomato	AT	423.46±6.31 <sup>Aa</sup>	239.69±3.78 <sup>ld</sup>	181.33±4.04 <sup>Cf</sup>	150.66±3.12 <sup>Dg</sup>	ND	ND	
	LT	423.46±6.31 <sup>Aa</sup>	371.92±5.07 <sup>Eb</sup>	287.84±3.20 <sup>Fe</sup>	247.16±3.17 <sup>lcd</sup>	209.46±2.70 <sup>Jh</sup>	169.28±2.42 <sup>Hi</sup>	

AT - Ambient temperature (32±2°C), LT - Lowl temperature (7±2°C)

ND - Not doneMean ± Standard Deviation values with different superscripts a, b, c, .... at p<0.01 & A, B, C, .... at p<0.05 differ significantly

AT and LT, respectively. There was no significant difference in % moisture content of the treated tomato from the corresponding untreated control after any days of storage at both the storage temperatures (Table 3).

**Titratable acidity as citric acid (g per 100 g dry weight):** Titratable acidity of the treated as well as non-treated tomatoes was  $7.63\pm0.24$  g per 100 g dry weight, which increased significantly (p<0.01) after 4 and 16 days of storage at AT and LT, respectively (Table 3). This was supported by the fact that tomato is a climacteric fruit (Burton, 1982), where respiration increases first and then decreases at later stages during ripening (Wills *et al.*, 1981). Organic acids too increase during storage of such fruit resulting in increased titratable acidity. It showed no significant (p<0.01) variation from that of the untreated control during storage for any days studied at both temperatures in case of the treated tomato (Table 2).

Ascorbic acid content (mg per 100 g dry weight): Ascorbic acid content of both the treated and the untreated control tomatoes decreased significantly (p<0.01) even after 4 days of storage at AT as well as LT (Table 4). There was no significant variation in the ascorbic acid content of the treated and the untreated tomatoes on any day of storage, carried out at both the temperatures, except 4 days of storage at AT, wherein the samples showed significant (p<0.01) difference in the ascorbic acid content (Table 4).

**Lycopene content (mg per 100 g dry weight):** Lycopene content of both the treated and the untreated tomatoes was  $34.41\pm3.22$  mg per 100 g dry weight, which increased significantly (p<0.01) after 4 and 8 days of storage at AT and LT, respectively (Table 4). The increase in the lycopene content during storage of the treated and the untreated tomatoes was significantly (p<0.01) greater at

AT than LT upto 8 days of storage, subsequent to which the difference was insignificant (Table 4). This finding was confirmed from the earlier studies, which showed that lycopene content of stored tomatoes at AT was higher than that at LT (Ajlouni *et al.*, 2001; Hamauzu *et al.*, 1998). The treated sample had non-significantly lower lycopene content than the untreated control during storage in most of the cases, except LT storage for 8 and 12 days, which was significant (p < 0.01) (Table 4).

Uptake of MBC residue (µg per g sample) was lowest in apple followed by banana, orange, tomato, okra, grape (golden) grape (blue black) and sapota. Low level of uptake of MBC residue in some fruits and vegetables was probably due to presence of waxy layer or characteristic features of peels. Dip treatment with MBC suspension was able to prolong to a greater extent the storage life of tomato than that of apple and at ambient temperature than that at chill temperature. The dip treatment of tomato decreased percent cumulative physiological loss in weight significantly during storage. Therefore, post-harvest controlled MBC treatment of selected fruits and vegetables would result in significant increase in their shelf-life at room temperature, and could be beneficial as the residues remain below tolerance limits.

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### References

Abu-Baker, F. and M.N.B. Abdul-Karim, 1994. Chemical treatments for microbial control on sapota. *ASEAN Food J.*, 9(1): 42-43.

- Agnihotri, N.P. 1999. Monitoring of pesticide residues. In: N. P. Agnihotri (ed.), *Pesticide Safety Evaluation and Monitoring*. All India Coordinated Research Project on Pesticide Residues, New Delhi, p. 9-23, 94-118.
- Ahuja, A.K., S. Mohapatra, D. Sharma and M.D. Awasthi, 1998. Monitoring of vegetables for pesticide contamination at harvest. In: *Proceedings of National Symposium Pest Management of Horticultural Crops*, Bangalore, p. 243-246.
- Ajlouni, S., S. Kremer and L. Masih, 2001. Lycopene content in hydroponic and non-hydroponic tomatoes during post-harvest storage. *Food Aust.*, 53(5): 195-196.
- Amadioha, A.C. 1996. Control of storage rot of potato caused by *Rhizopus oryzae. Int. J. Pest Manag.*, 52(4): 311-314.
- Amadioha, A.C. 1998. Control of post harvest tuber root of potato incited by *Rhizoctonia bataticola*. Arch. Phytopath. Plant Protec., 31(3): 225-231.
- AOAC, 1990. Official Methods of Analysis. Association of Official Analytical Chemists, 15 th edn, Vol 2, AOAC Inc, Arlington, Virginia, 22201, USA, pp 912, 918, 1058.
- Awasthi, M.D. 1998. Pesticide residues in food chain in India fruits and vegetables. In: O. P. Shukla and A. K. Omkar Kulshrestha (eds.), *Pesticides, Man and Biosphere*. APH Publishing Corporation, New Delhi. p. 121-160.
- Awasthi, M.D. and A.K. Ahuja, 1997. Occurrence of pesticide residues in market and farm gate samples of vegetables in and around Bangalore. J. Food Sci. Technol., 34: 146-149.
- Awasthi, M.D. and D. Sharma, 1997. Uptake of fungicide residues and their persistence on ripening mango fruits from post-harvest treatments. *Pesti. Res. J.*, 9: 41-45.
- Baldi, M., L. Zanoni and A. Bovolenta, 1981. Benzimidazole fungicide contamination of apples, pears and strawberries from Ferrara province. *Indust. Alim.*, 20(9): 594-595.
- Baldi, M., S. Maietti and M.C. Pietrogrande, 1982. Simultaneous determination of residues of ethoxyquin, TBZ, and MBC by a combined technique. *Indust. Alim.*, 21(199): 771-776.
- Baur, P., H. Marzouk, J. Schonherr and B.T. Grayson, 1997. Partition coefficients between plant cuticle and adjuvants as related to foliar uptake. J. Agric. Food Chem., 45: 3659-3665.
- Beerh, O.P. and G.S. Siddappa, 1959. A rapid spectrophotometric method for the detection and estimation of adulterants tomato ketchup. *Food Technol. (Chicago)*, 13: 414-418.
- Bencivenga, B., G. Pallotti, G. Pasquazi, I. Rosatelli and T. Simonetti, 1982. Residues of dithiocarbamate and benzimidazole pesticides in fruits and vegetables in the Rome Market. *Indust. Alim.*, 21(198): 687-689.
- Ben-Yashoshua, S. and E. Cohen, 1981. Decay control and fungicide residues in citrus fruits seal packed in HDPE. *Pesti. Sci.*, 12(5): 485-490.
- Burton, W.G. 1982. *Post-harvest physiology of food crops*, Longman, London, pp 55.
- Cabras, P., A. Angioni, V.L. Garan, M. Melis, F.M. Pirisi, F. Cabitza and M. Cubeddu, 1998. Pesticide residues on field-sprayed apricots and in apricot drying process. J. Agric. Food Chem., 46: 2306-2308.
- Cano, P. and J.L. Plaza, 1987. Determination and persistence of several fungicides in post-harvest treated apples during cold storage. J. Agric. Food Chem., 35(1): 144-147.
- Charles, R.W. 1987. *The Pesticide Manual*. 18th edition, 58-59, 127-128.
- Goulart, B.L., P.E. Hammer, K.B. Evensen, W. Janisiewicz and F. Takeda, 1992. Pyrrolnitrin, captan + benomyl, and high CO<sub>2</sub> enhance raspberry shelf life at 0 or 18 C. J. Amer. Soc. Hort. Sci., 117(2): 265-270.
- Gupta, V.K. and D. Mukherjee, 1980. Prolonging life of Allahabadi Safida guava with morphactin and benomyl. *Ind. J. Hort.*, 37(2): 163-166.

- Hamauzu, Y., K. Chachin and Y. Ueda, 1998. Effect of post harvest storage temperature on the conversion of <sup>14</sup>C-mevalonic acid to carotenes in tomato fruit. J. Jap. Soc. Hort., 67(4): 549-555.
- Hamil, S. and D.B. Harper, 1982. Carbendazim and ethoxyquin residues in apples and apple juice from Northern Ireland. *Rec. Agric. Res.*, 30: 33-37.
- Hargreaves, P.A. 1983. Benomyl residues in litchi after post-harvest dipping. Aust. J. Exp. Agric. Anim. Husb., 23(12): 95-98.
- Jahangir, Sher Hassan and Ahmad Shabir, 1994. Chemical control of post-harvest apple fruit rot (*Penicillum expansum*). Sarhad J. Agric., 10(3): 327-330.
- Kiigemagi, U., R.D. Inman, W.M. Mellenthin and M.L. Deinzor, 1991. Residues of benomyl (determined as carbendazim) and captan in post-harvest treated pears in cold storage. J. Agric. Food Chem., 39(2): 400-423.
- Marzouk, H., P. Baur and J. Schonherr, 1998. Relative solubilities of bigenox and 1-naphthyl acetic acid in plant cuticles and in selected pure or aqueous glycol additives. *Pesti. Sci.*, 53: 278-284.
- Meloni, M. and F.M. Pirisi, 1984. Residues of fungicides on greenhouse lettuce. J. Agric. Food Chem., 32(2): 183-185.
- Mohammed, M. and L. Sealy, 1988. Hydrocooling and post-harvest quality in melongene (*Solanum melongena* L.). *Tropi. Agric.*, 65(2): 161-165.
- Monico-Pifarrae, A. 1987. Monitoring residues of carbendazim (applied as benomyl) and thiabendazole in Wellspur apples. J.A.O.A.C., 70(3): 596-598.
- Nagayama, T., Y. Tamura and T. Maki, 1983. Survey of pesticide residues in imported teas and imported bananas. *Ann. Rep. Tok. Metro. Res. Lab. Publ. Healt.*, 34: 165-167.
- Nwufo, M.I., K.I. Okonkwo and J.C. Obiefuna, 1994. Effect of postharvest treatments on the storage life of avocado pear (*Persea* americana Mill.). Trop. Sci., 34(4): 363-370.
- Papadopoulou-Mourkidou, E. 1991. Post-harvest applied agrochemicals and their residues in fresh fruits and vegetables. J.A.O.A.C., 74(5): 745-765.
- Rangaswamy, J.R., Y.N. Vijayashankar and S.R. Prakash, 1987. Colorimetric method for the determination of carbendazim (MBC), benomyl and their degradative product-2-aminobenzimidazole. J Food Sci. Technol. (India), 24: 309-311.
- Rathu, A.S, A.S. Chharia and Ranjit Kumar, 1989. Effect of different fungicides on shelf life and quality of beauty seedless grapes. *Int. J. Trop. Agric.*, 7(3/4): 256-261.
- Sanderson, P.G. 1999 Fungicidal drenches for control of post-harvest decay. Good Fruit Grower, 50(5): 53-58.
- Sharma, R.C., J.L. Kaul and R.L. Sharma, 1989. Effect of post-inoculation fungicidal dips on post-harvest diseases of peaches. *Int. J. Trop. Plant Dis.*, 7(2): 221-224.
- Snedecor, G.W. and W.G. Cochran, 1989. *Statistical Methods*. Iowa State University Press, Ames, Iowa, USA.
- USDA, 1993. Agricultural Marketing Service, Pesticide Data Programme Jan-June 1992 Report. US Department of Agriculture, Washington DC.
- Wasker, D.P., P.B. Khedkar and V.K. Garande, 1999a. Effect of postharvest treatments on storage behaviour of pomegranate fruits under room temperature and cool storage. *Ind. Food Pack.*, 53(2): 5-9.
- Wasker, D.P., P.B. Khedkar and V.K. Garande, 1999b. Effect of postharvest treatments on shelf life and quality of pomegranate in evaporative cool chamber and ambient conditions. J. Food Sci. Technol. (India), 36: 114-117.
- Wills, R.H.H., T.H. Lee, D. Graham, W.B. McGlasson and E.G. Hall, 1981. Post-harvest. New South Wales University Press, Kensington.